quercetin, kampferol and cyanidin (from leuco-cyanidin) have been found in the hydrolysed leaf extract of *C. reliaiosum* [1].

Present work. Extraction and identification of the flavonoids from the bark. The crude flavonoids were obtained by extraction of the milled bark with ethanol in the usual manner [2] and purified by means of column chromatography and preparative TLC. Apigenin (5,7,4'-trihydroxyflavanone), naringenin (5,7,4'-trihydroxyflavanone) and (+)-afzelechin (5,7,4'-trihydroxyflavan-3-ol) were identified by NMR, IR, mmp and CoTLC and derivative formation [3–6]. The glycosides prunin (naringenin 7-O-glucoside) and cosmosiin (apigenin 7-O-glucoside) were identified by their physical data [6,7] and hydrolytic conversion to glucose together with naringenin and apigenin respectively.

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# ACHILLIN AND DEACETYLMATRICARIN FROM TWO ARTEMISIA SPECIES

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**Key Word Index**—Artemisia ludoviciana; A. klotzchiana; Compositae; essential oil; camphor; borneol; achillin; deacetylmatricarin; 3,5-dihydroxy-6,7.8-trimethoxyflavone.

Plants. Artemisia ludoviciana, Nutt (Voucher specimen 7294) collected in Punta de la Loma, N.L.; A. klotzchiana, Basser (Voucher specimen 7393) collected in San Roberto, N.L. Both plants have the trivial name of "estafiate" and are used for stomach ailments.

Previous work. The Artemisia genus contains more than 300 species and geographical races many of which have been scrutinized by chemists [1]. Four santanolides, ludovicin-A, -B, -C [2] and ludolbin [3] has been isolated from A. ludoviciana, but only the essential oil of A. klotz-chiana has been studied, GLC showing principally camphor and borneol.

Present work. The sesquiterpene lactones, deacetylmatricarin and achillin, originally obtained from other members of the same tribe the Anthemideae, have been found in both species and supports the concept of geographic races. The high

yield of camphor and borneol from the essential oil of A. ludoviciana may have an important economical and chemotaxonomic value. The presence of 3,5-dihydroxy-6,7,8-trimethoxy flavone, an isomer of euparolin and eupatilin the cytotoxic flavonoids from Eupatorium serratum [5], also is of chemotaxonomic interest.

### **EXPERIMENTAL**

Steam distillation. From 1 kg of the aerial part of fresh A. ludoviciana, 12 ml of yellowish oil were obtained, sp. gr. $_4^{4.5}$  0-897;  $\eta_D^{2.5}$  1-4764 [ $\alpha$ ] $_0^{2.5}$  + 2; GLC showed camphor, (52%) borneol (25%), phellandrene (4%),  $\alpha$ -pinene (3%), plus 13 other minor constituents. As reported, the aerial part (1 kg) of A. klotzchiana afforded 8 ml of a bluish essential oil  $\eta_D^{2.5}$  1-4665 [ $\alpha$ ] $_0^{2.5}$ , rich in camphor (35%) and borneol (40%).

Extraction of dried and milled plant material. Light petrol extraction of 498 g of A. ludoviciana afforded 1·22 g of achillin, mmp Co-TLC IR, NMR,  $[\alpha]$  together with 0·84 g of camphor. A. klotzchiana (500 g) was extracted with CHCl<sub>3</sub> affording 15 g of resinous extract, which on Si gel chromatography gave 506 mg of deacetyl-matricarin mmp Co-TLC, IR, NMR,  $[\alpha]_D^{2.5}$ 

and 57 mg of 3,5-dihydroxy-6,7.8-trimethoxyflavone, yellow plates mp 228–229°  $C_{17}H_{14}O_{7}$  (M $^+$  344) IR, 3400, 3030, 1650, 1570, 1550, 1490, 1450, 1400, 1350, 1310, 1290, 1250, 1200, 1140, 1100, 1090, 1030, 1010, 980, 950, 890, 850, 820, 780 cm $^{-1}$ ; chemical tests [6], MS, NMR (CDCl $_{31}$ TMS, in  $\delta$ ), 8·05 d (1H, 1Hz), 7·90 d (1H, 1Hz), 7·75 d (1H, 1Hz), 7·35 d (1H, 1Hz), d (1H), 4·10 (s, 6H), 4·0 (s, 3H), UV  $d_{\rm max}^{\rm MSOH}$ , 242 nm (d 23243), 275 (22726), 340 (30991), bathochromic shifts of band I with AlCl $_{3}$ , NaOMe, NaOAc as expected [7]. KOH fusion gave benzoic acid (TLC, PC and mmp).

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# PYRAZOLE IN CITRULLIS VULGARIS (CUCURBITACAE)\*

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Key Word Index—Citrullis vulgaris; Cucurbitacae; pyrazole; spectrophotometry.

L- $\beta$ -Pyrazolyl-alanine and  $\gamma$ -L-glutamyl- $\beta$ -pyrazolyl-L-alanine are found in seeds of many species of the Cucurbitaceae [1]. The former is synthesized by watermelon seedling extracts from pyrazole and O-acetylserine [2], while extracts of several other cucurbits were earlier reported to synthesize the compound from pyrazole and serine [3]; presumably, in these cases, the crude extracts could convert the amino acid to its Oacetyl derivative. A C<sub>6</sub>H<sub>6</sub> extract of ground cucumber seeds heated with serine, pyridoxal phosphate and aluminum sulfate was shown to vield  $\beta$ -pyrazolyl-alanine [3], providing indirect evidence that pyrazole existed in the seed. However, the presence of free pyrazole was never confirmed nor its concentration determined at the time [3] for lack of a sensitive and specific assay.

In the presence of a mild oxidant, pyrazole forms a yellow complex with trisodium pentacyano-aminoferrate (TPF). This property has been applied to the quantitative analysis of pyrazole in microbial broths [4] and in blood [5]. We found that the TPF reagent could be used to demonstrate the presence of pyrazole in

extracts of seed of Citrullus vulgaris (watermelon, var Coles Early and Early Canada). However, pyrazole was not detected in seed extracts of five varieties of squash (Cucurbita melopepo), 2 varieties of pumpkin (Cucurbita pepo), two varieties of cantaloupe (Cucumis melo), two varieties of cucumber (Cucumis sativus) and one variety of citron (Citrullus vulgaris var citroides).

The quantitative determination of pyrazole in watermelon seeds indicated a concentration of 410  $\mu$ g/g in var Early Canada and 280  $\mu$ g/g in var Coles Early. In 6-day old seedlings of var Early Canada, the concentration was 60  $\mu$ g/g fr. wt in the root and 140  $\mu$ g/g fr. wt in the hypocotyl.

Organic compounds containing the N-N bond have been reported in bacteria and fungi. Their presence in higher plants [1,6] is apparently rare, though this may reflect a lack of tests suitable for their detection. How plants synthesize or degrade the N-N bond remains a fascinating question for future study.

### **EXPERIMENTAL**

Preparation of extracts. Seeds or other tissues were ground and extracted for 24 hr with 65°, EtOH (3 to 5 ml/g) and extracts clarified by centrifugation at 3000 a.

<sup>\*</sup> NRCC No. 14917.